



**UNIMORE**

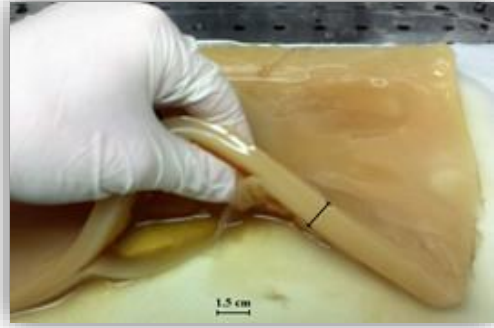
UNIVERSITÀ DEGLI STUDI DI  
MODENA E REGGIO EMILIA

# Application of versatile Acetic Acid Bacteria to innovative bioprocesses

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Supervisor: Dr. Maria Gullo

Ph.D. Workshop in Agri-Food Sciences, Technologies and Bio-Technologies  
1 December 2017 , Reggio Emilia (RE) Italy

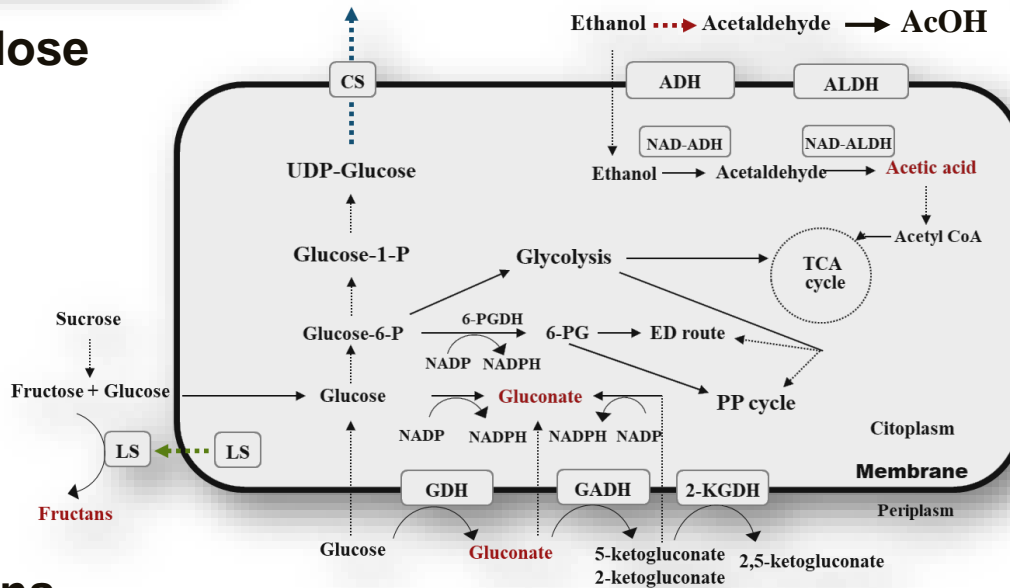
# Versatility of Acetic Acid Bacteria



Cellulose



New vinegars



Bioactive attractants

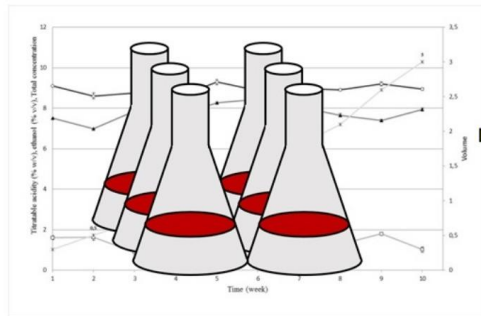
Fructans



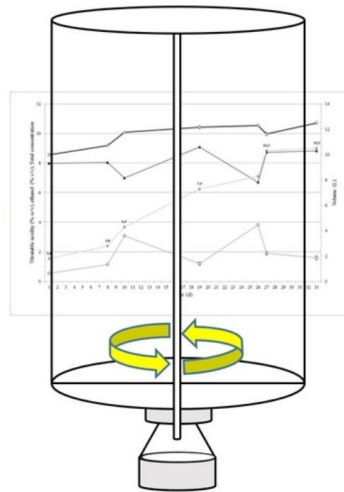
# Part 1

## Feasible acetic acid fermentations of alcoholic and sugary substrates in combined operation mode

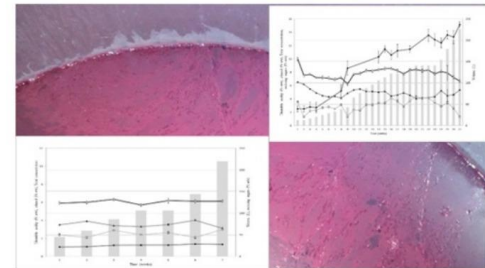
**Microscale cultivation  
in static system**



**Submerged  
fermentation**



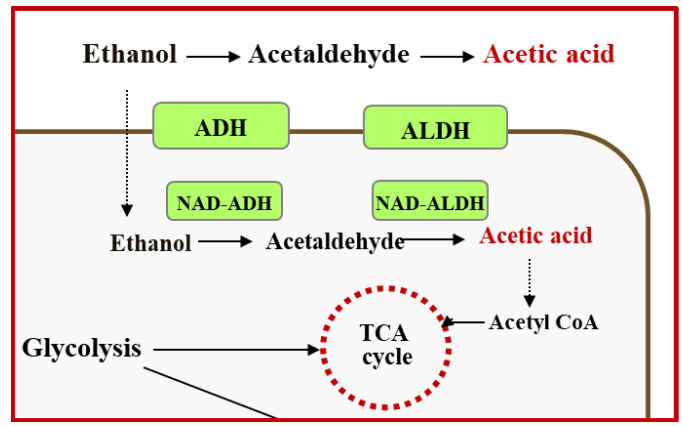
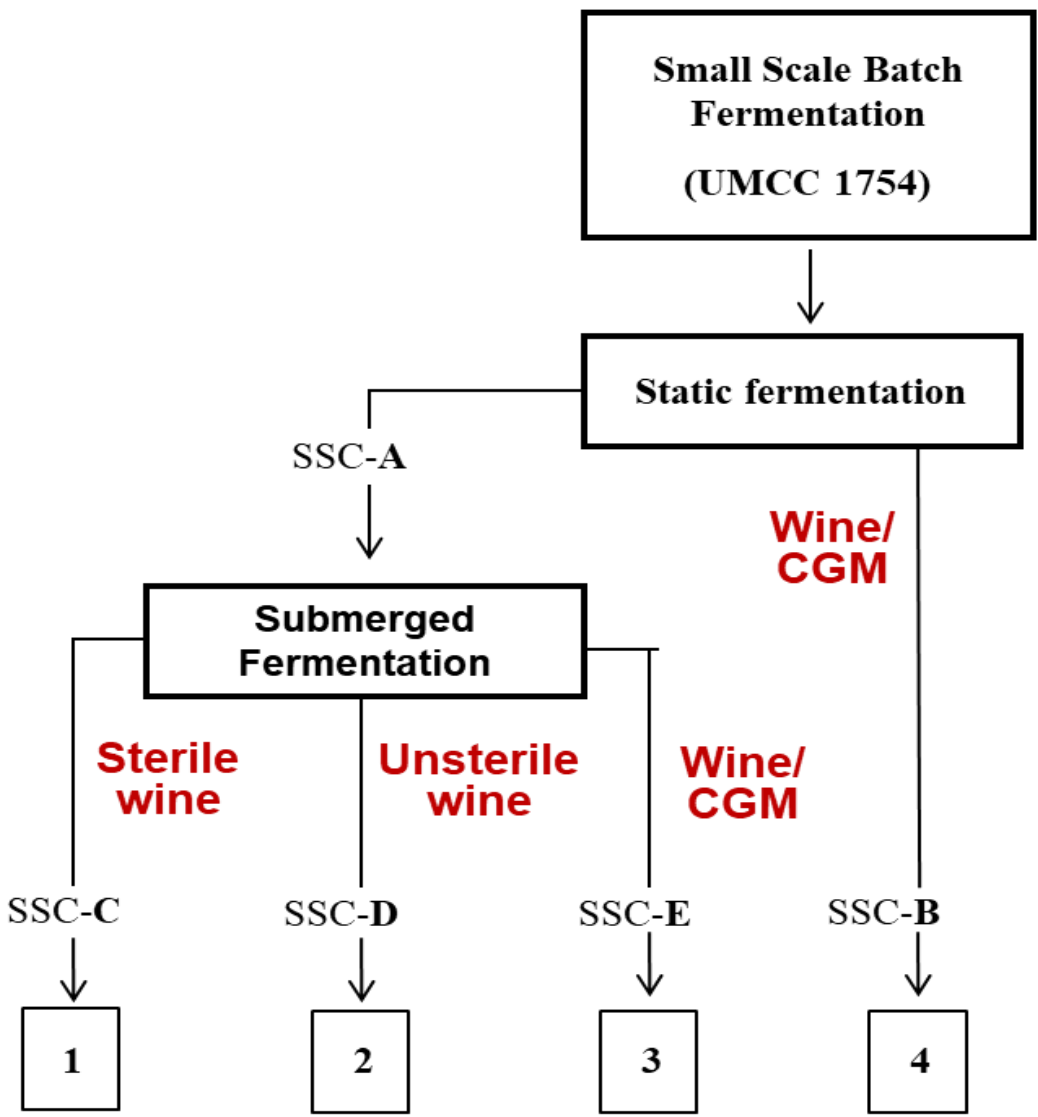
**Prototypal scale  
fermentation**



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Process Biochemistry 51, 1129-1139, 2016

# Part 1 Implementation of a combined system (static and submerged)



## A. pasteurianus UMCC 1754

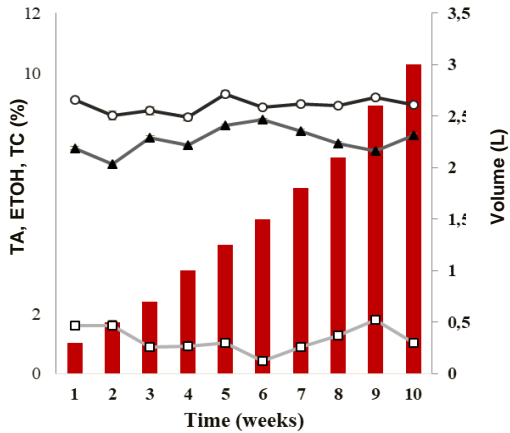
- High acetic acid production rate
- Efficient start-up and persistence
- Phenotypic stability
- No production of undesired products
- Low nutritional needs

**4 DIFFERENT VINEGARS**

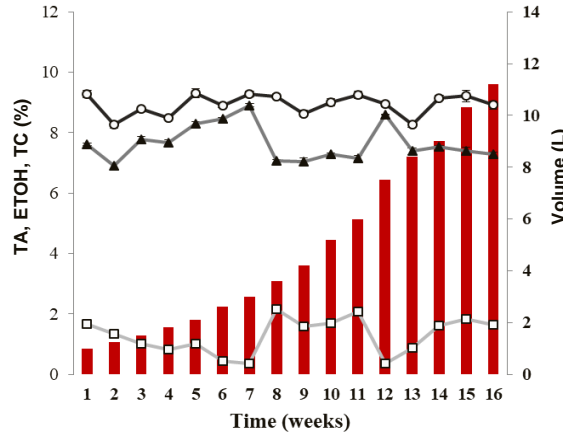
**STATIC SYSTEM**

TA: Titratable acidity; EtOH: Ethanol; TC: Total concentration

**SSC-A**

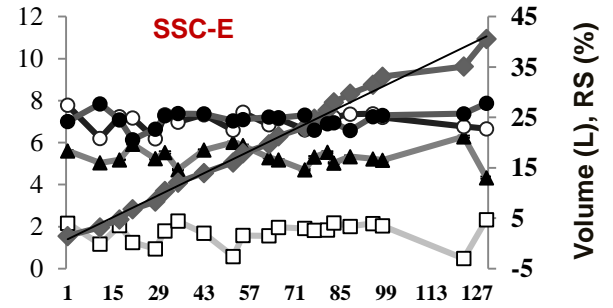
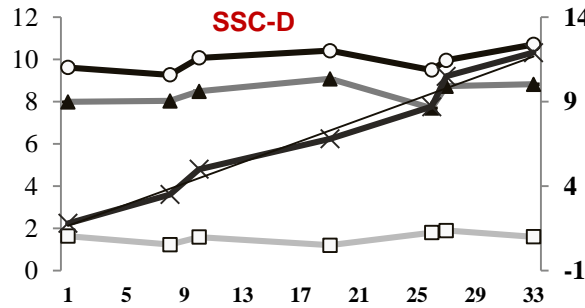
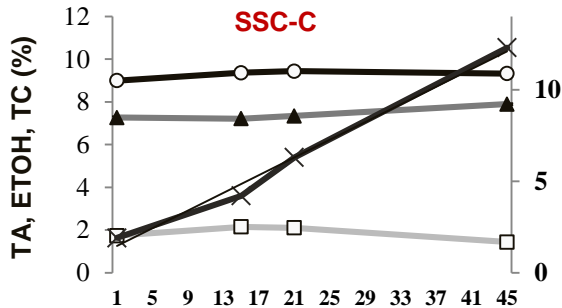


**SSC-B**



- SSC-A to start-up submerged fermentation
- SSC-B to prototype scale
- Slow fermentation
- SSC-C, D wine
- SSC E grape must
- Fast fermentation

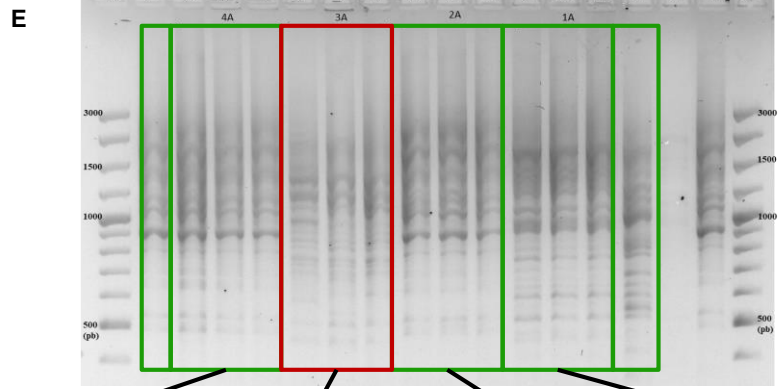
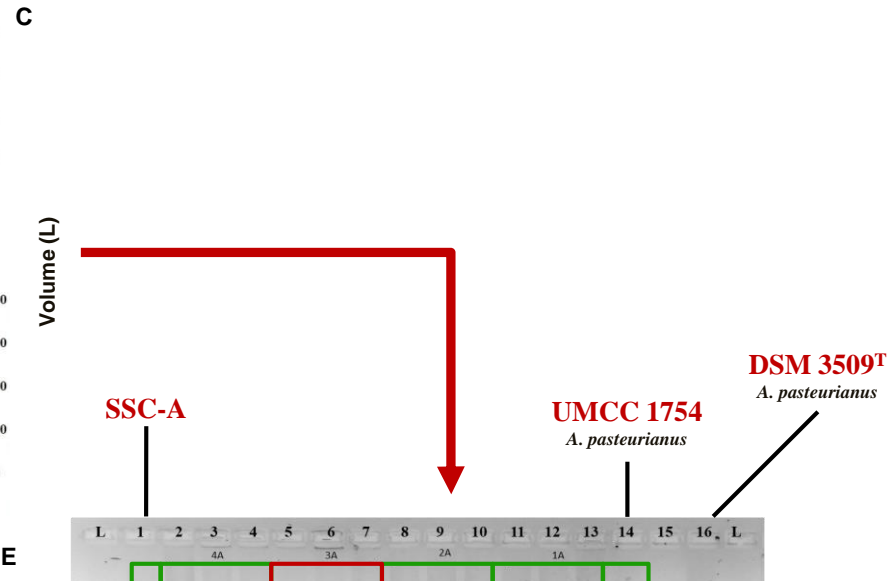
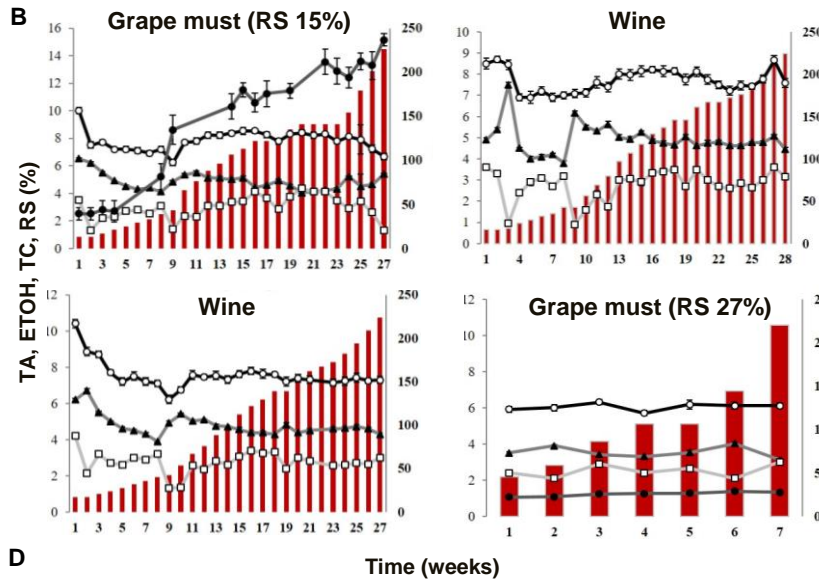
**SUBMERGED SYSTEM**



Time (Days)

# Part 1

# Cultures performance at prototypal scale

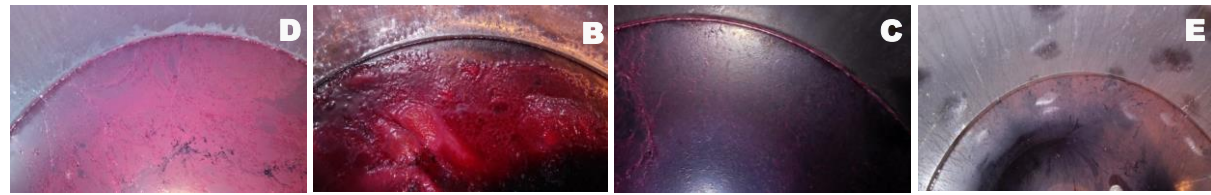


## SCALE-UP

Over 200L of vinegar/batch in 6 months

**BATCH B-E: Wine**

**BATCH C-D: CGM**



## Conclusions

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**Combined fermentation by *A. pasteurianus* UMCC 1754 yielded viable SSCs at both laboratory and prototype scales**

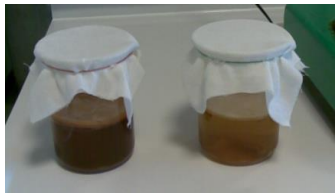


**Process stability in static, submerged and prototype-scale confirmed the feasibility of using SSCs in industrial vinegar fermentations**

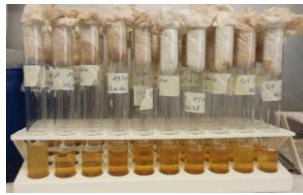


# Part 2

## Increased production of bacterial cellulose as starting point for scaled-up applications



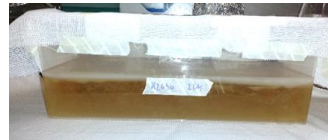
ISOLATION



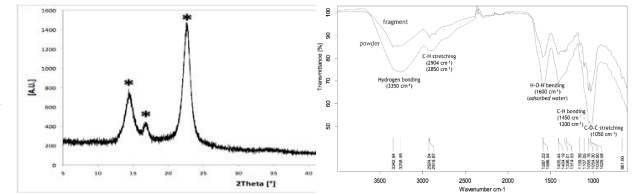
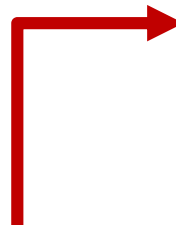
QUALITATIVE TEST



QUANTITATIVE TEST



MEDIUM OPTIMIZATION



STRUCTURAL CHARACTERIZATION



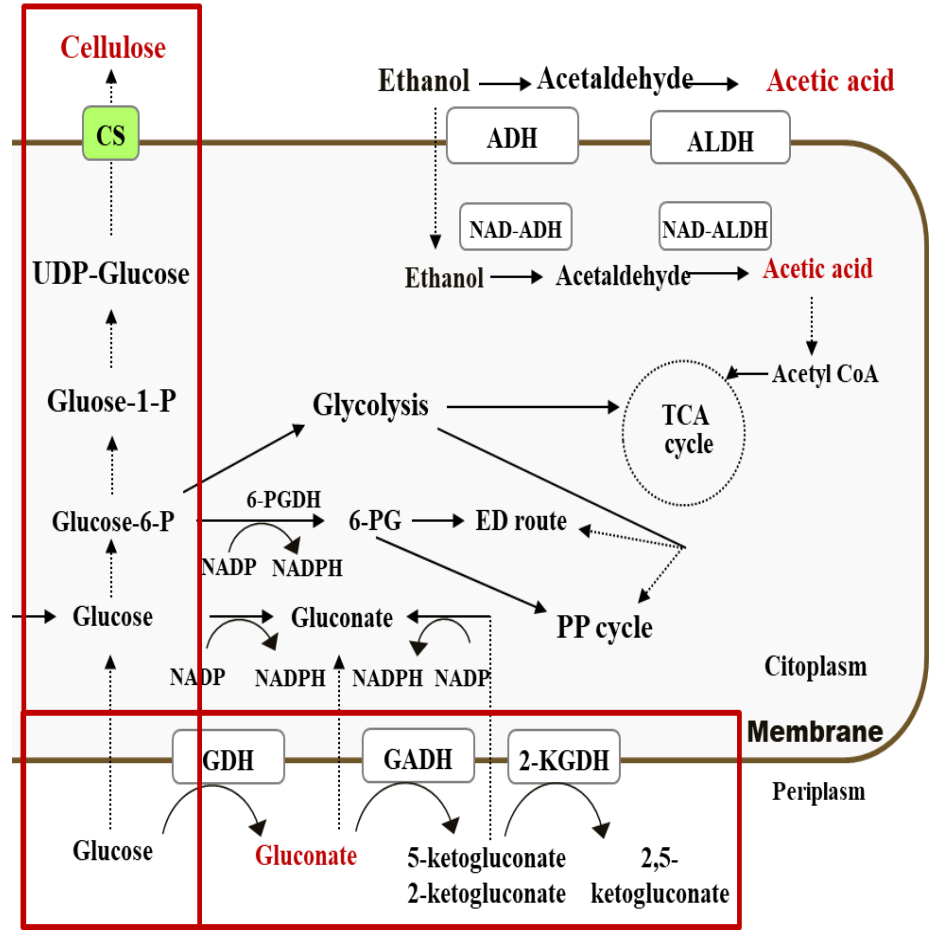
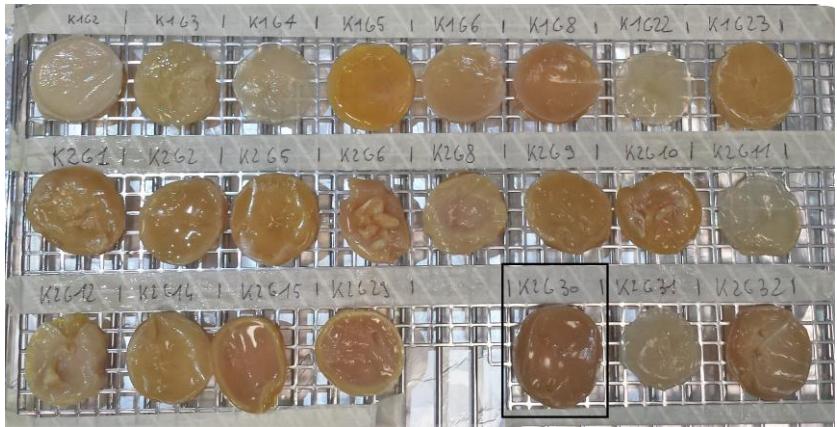
Published on:

Applied Microbiology and Biotechnology 101, 2017



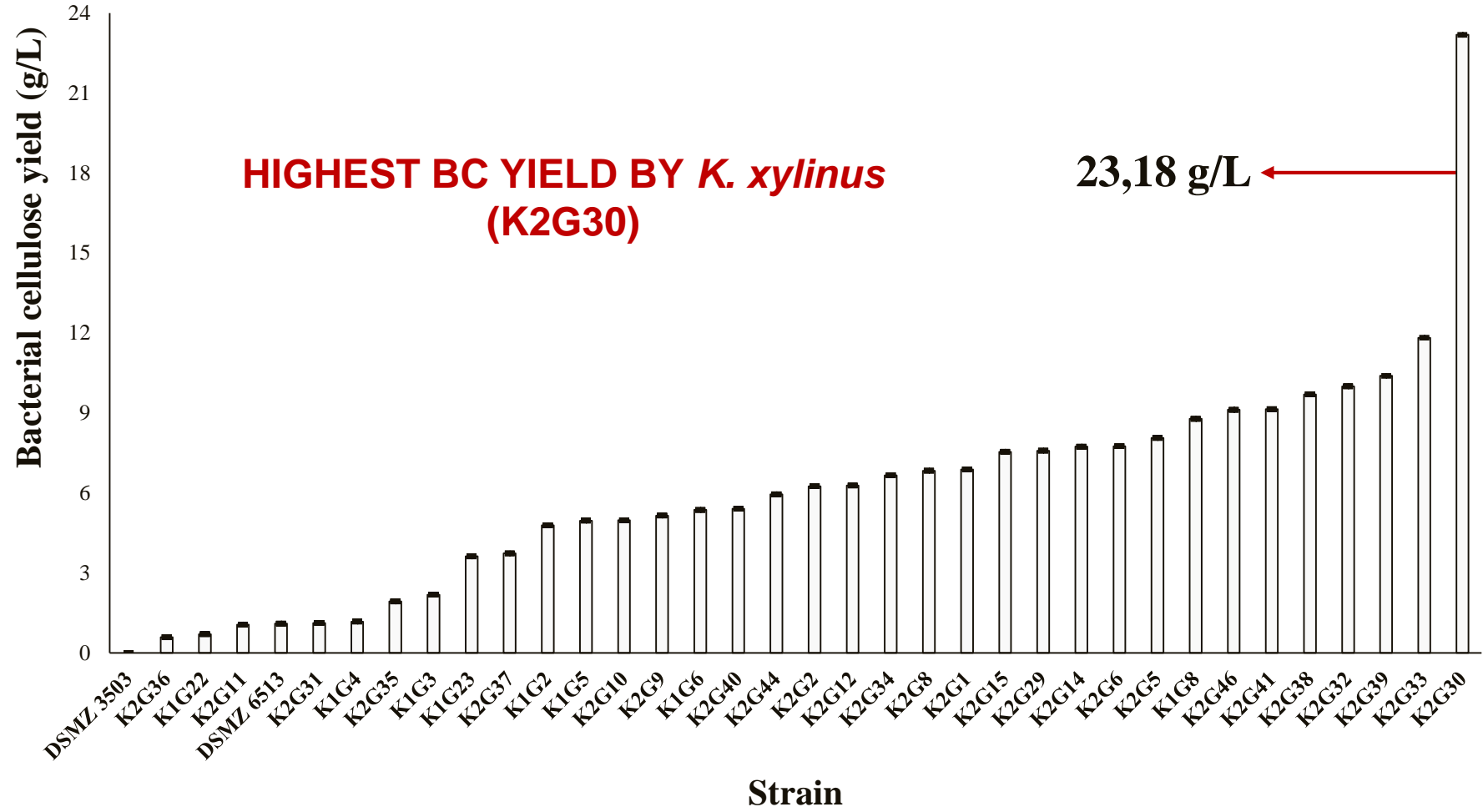
# Bacterial cellulose synthesis

- D-glucose with  $\beta(1-4)$  glycosidic bond
- Synthesized by cellulose synthase (CS)
- High purity
- High degree of crystallinity
- High W.A.R. and resistance to tensile strength



# Cellulose production

**34 STRAINS SHOWED BC PRODUCTION**



# Culture optimization

Strain K2G30

UMCC 2756 (*K. xylinus*)



**FORMULATION OF AN ENHANCED BROTH**

**GET = GY- CaCO<sub>3</sub>+ 1.4% ETOH**

**+**

**S/V RATIO OPTIMIZATION**

**0,23 cm<sup>-1</sup>**



**HIGHER BC YIELD**

**LOWER BY-PRODUCTS FORMATION**

**Ethanol is an additional energy source so it allows glucose to be used mainly for BC synthesis**

# CARBON SOURCES CONSUMPTION AND BC PRODUCTION

Part 2

15 days of static cultivation in vessel (S/V 0.23 cm<sup>-1</sup>)

Broth	pH		Glu (g/L)		*EtOH (g/L)	G.A (g/L)	A.A (g/L)	BC g/L	BC/ cons sugar (g/g)
	Initial	final	initial	final	final				
<b>GY</b>	5.40	3.14	50.00 ± 0.03	15.73 ± 0.03	0.00	22.23 ± 0.05	0.00	13.25 ± 1.11	0,39
<b>GET</b>	6.37	4.45	50.00 ± 0.02	18.00 ± 0.08	2.20 ± 0.08	11.46 ± 0.05	0.67 ± 0.08	19.64 ± 0.94	0,61

## OPTIMIZED CONDITIONS

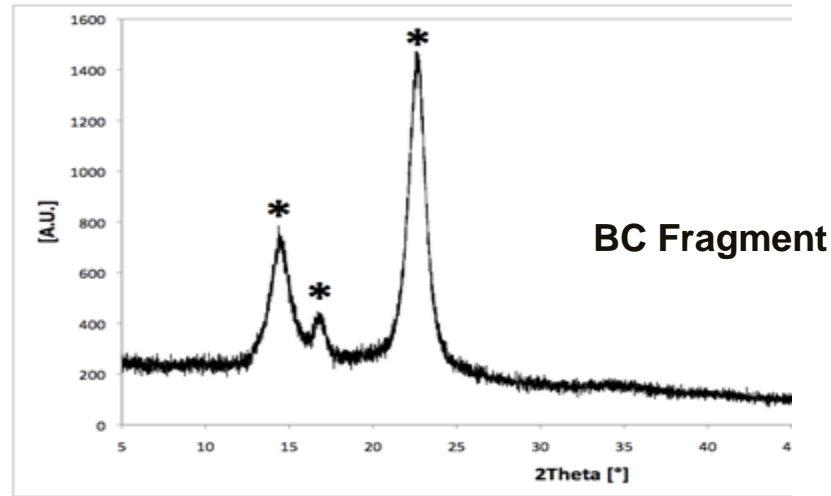
- BC production increased of 30%
- Gluconic acid was ~ 50% lower
- Final pH was higher
- g of BC produced per consumed sugar increased

\*GET: Initial Ethanol: 14%  
**Glu:** glucose  
**EtOH:** ethanol  
**GA:** gluconic acid  
**A.A:** acetic acid  
**BC:** bacterial cellulose

## Part 2

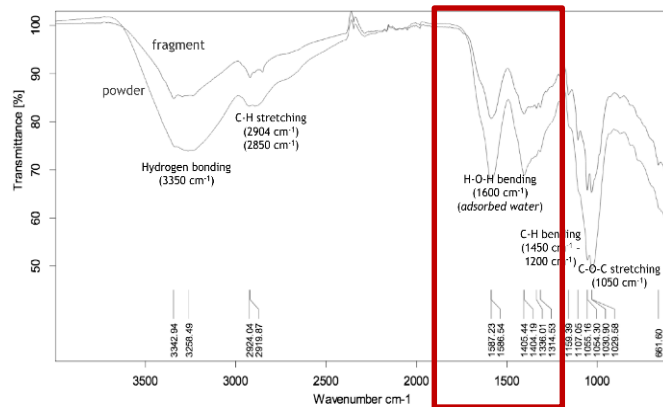
# Structural analysis of bacterial cellulose produced by K2G30

## XRD



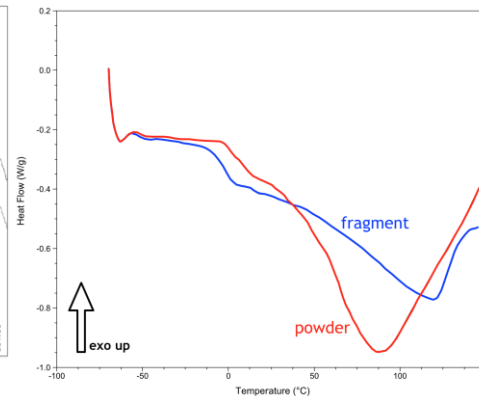
- Typical peaks of BC with high degree of crystallinity
- **D.o.c = > 80%**
- Graphs are analogous and the peaks are typical for BC.

## FT-IR



**Absorbed water**

## DSC



- **Chemical bonds were not altered by the milling process**
- **Glass transition temperature (T<sub>g</sub>) at about 0 °C.**
- **The mass loss (water removal) was 12.9%.**
- **W.A.R: 400%**

## Conclusions

- The highest yield was achieved by K2G30 by static cultivation among 34 studied strains
- Enhanced culture conditions increased BC production of 30%
- High-purity BC with high degree of crystallinity (80%) and WAR (400%)



**Selected strain was able to produce high amount of BC suitable for biomedical applications and food processing**

## Part 3

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# Screening and selection of acetic acid bacteria for fructans production



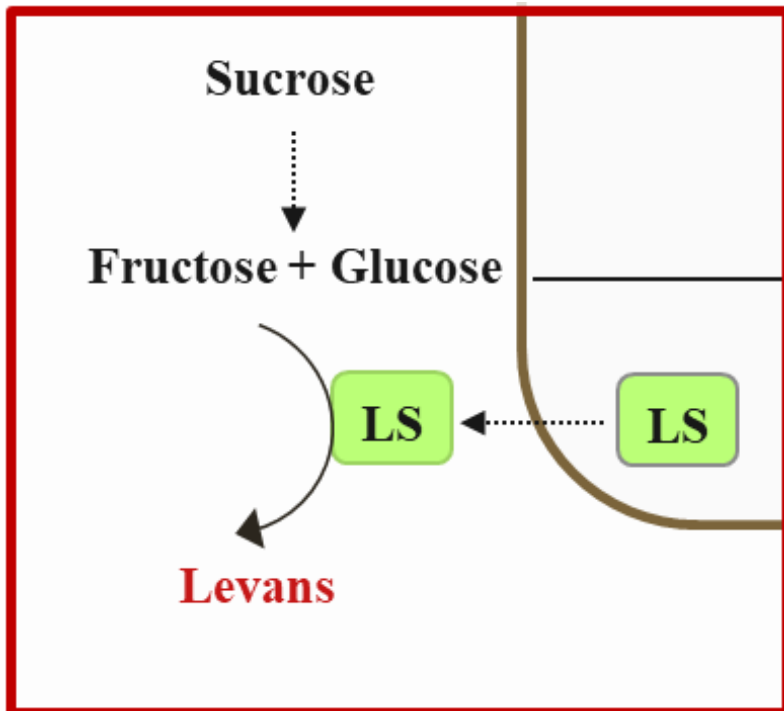
# Background, aim and strategy

- Some AAB are levan producers  $\beta(2\rightarrow6)$
- Prebiotics, anti-cancer and anti-viral effect
- Thickening and gelling agents
- Biodegradable plastics, glues, cosmetics, textile coatings, detergents

Some AAB are high fructan (Levan) producers



**Selection of strains to develop prebiotic beverages**



REVITALIZATION



FRUCTAN TEST

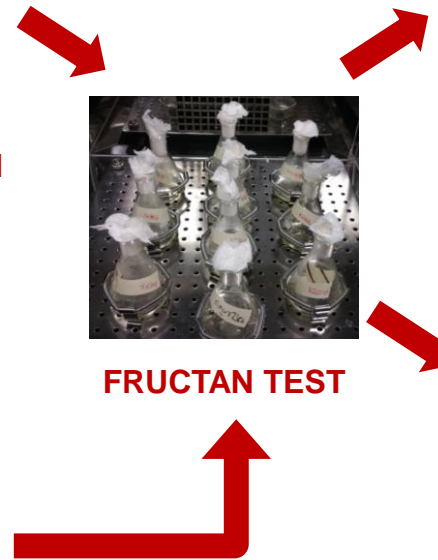


ASSAYS



EXTRACTION AND CHARACTERIZATION

MEDIUM OPTIMIZATION



## Identification of the screening conditions

Strain	Sucrose concentration	
	250 g/L	300 g/L
NBRC 101099 <sup>T</sup>	+	+
UMCC1754	+	weak
UMCC 1789	+	weak
DSM 3509 <sup>T</sup>	+	+
DSM 2343	+	+
DSM 2004 <sup>T</sup>	+	+
UMCC 2756	+	+



### OSMOTOLERANCE SCREENING

- Most AAB strains can grow at 300 g/L of sucrose
- *A. pasteurianus* strains prefer lower concentration

**250 g/L MAXIMUM SUCROSE CONCENTRATION**

## SCREENING OF FRUCTAN PRODUCTION

### STANDARD CONDITIONS

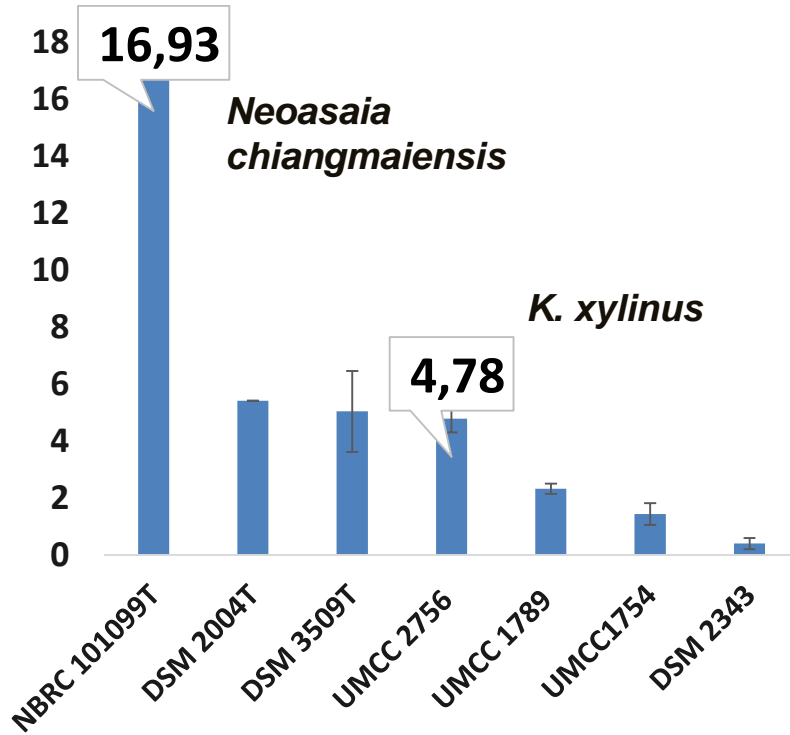
- Sucrose concentration: 70 g/L
- Shaking speed: 140 rpm

### OPTIMIZED CONDITIONS

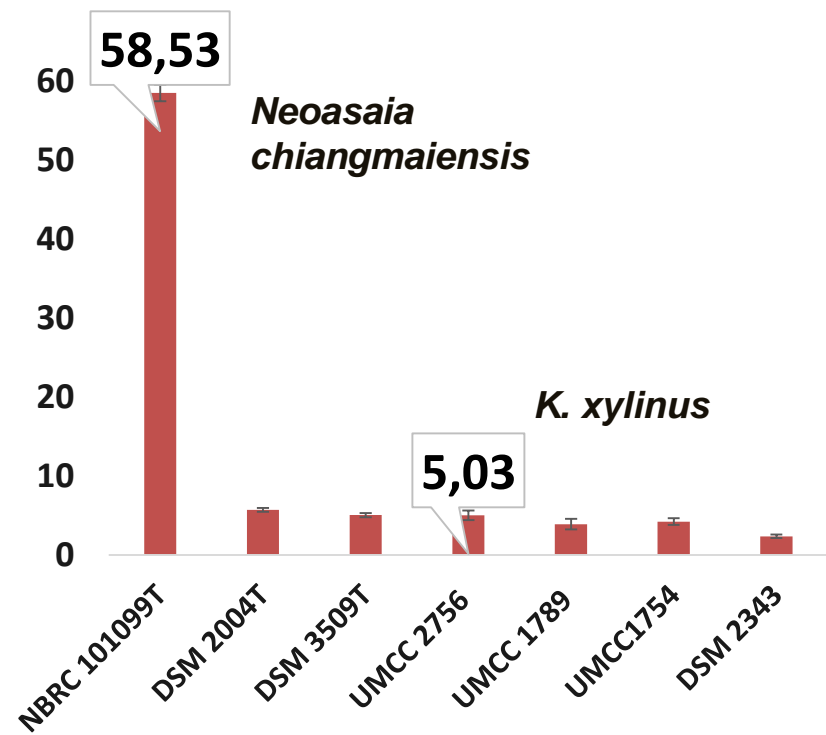
- Sucrose concentration: **250 g/L**
- Shaking speed: **200 rpm**

# Fructan production optimization

## STANDARD CONDITIONS

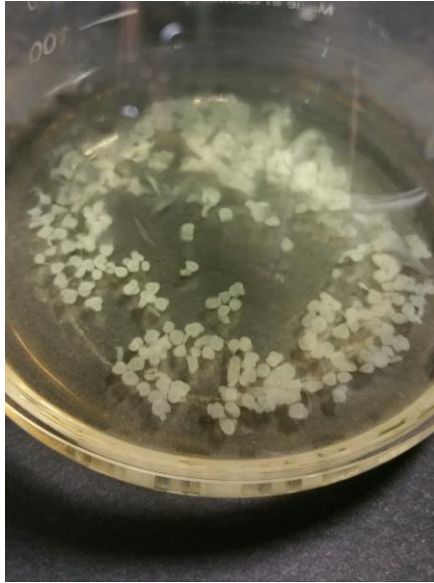


## OPTIMIZED CONDITIONS



- *Neosasaia chiangmaiensis* showed the highest fructan production in both conditions
- *Acetobacter* and *K. xylinus* strains produced about 5 g/L in both conditions

# CELLULOSE SCREENING AND EXTRACTION



Strain	QUALITATIVE BC PRODUCTION TEST	
	HS-G	HS-S
NBRC 101099 <sup>T</sup>	-	-
DSM 2343	-	-
DSM 2004 <sup>T</sup>	+	+
DSM 3509 <sup>T</sup>	-	-
UMCC1754	-	-
UMCC 1789	-	-
UMCC 2756	+	+



## FRUCTANS EXTRACTION PROTOCOL

- Centrifugation
- + 2 Volume of cold ETOH
- Precipitation
- Centrifugation
- Levan-pellets collected
- Levan purification



## FRUCTAN CHARACTERIZATION

Preliminary results showed a FT-IR spectra of pure levan

Fructans extracted by ethanol precipitation



## Conclusions

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***Neosasaia chiangmaiensis* NBRC101099 showed the highest fructan production in both standard and optimized culture conditions**



**Strains suitable for industrial production**

***Komagataeibacter* strains showed a considerable production of both fructans and cellulose**



***K. xylinus* strain UMCC 2756 is suitable to develop functional foods and beverages**

# General conclusions of the Ph.D. project

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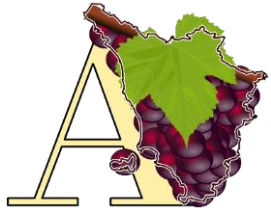
- **High versatility of Acetic Acid Bacteria opens wide perspectives in both food and non food industry**
- **Possibility to develop new products**
- **Research on small scale is essential to improve the processes on large scale**

**Performing a further engineering of the investigated processes is the most suggested development, both in research and industrial scale**

# A special thanks to...



## Vignola Foundation (Modena, Italy)



Aceti speciali da mosti d'uva della toscana.  
MISURA 124-PSR 2007-2013  
Regione Toscana

Saporea, Verona, Italy

San Giacomo s.r.l

Antonella Sola of



Monia Montorsi of

**UNIMORE**

Dipartimento di Scienze  
e Metodi dell'Ingegneria